

1 Management of scleractinian coral assemblages in temperate non-reefal areas: Insights
2 from a long-term monitoring study in Kushimoto, Japan (33°N)

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25

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36 **Abstract:** In this era of global climate change, understanding fundamental mechanisms
37 of coral community maintenance and persistence in temperate non-reefal areas is a high
38 marine conservation priority. To identify mechanisms of community maintenance and
39 persistence via larval supply, we monitored coral settlement over 12 years and
40 investigated the genetic population structure of two major acroporid species at Kushimoto,
41 Wakayama Prefecture, Japan (33° N). From 8 to 30 artificial settlement panel pairs were
42 deployed from May or June to September, October, or November of each year. Recruits
43 on settlement panel pairs were scarce, especially those of acroporids (0 or < 1 recruit per
44 panel pair in most years). As coral cover in the Kushimoto area remained relatively high
45 over a decade, such low recruitment may be sufficient for persistence of acroporid
46 communities in this region. In addition, genetic analysis using 8 or 10 microsatellite
47 markers demonstrated differences in genetic structure between populations of *Acropora*
48 *hyacinthus*, which is a long-term resident species in this area, and *A. muricata*, a recently
49 arrived species. *Acropora hyacinthus* displayed higher numbers of multilocus genotypes
50 (41 of 43 samples collected) whereas only one multilocus genotype in 30 samples was
51 seen in *A. muricata*. This difference may reflect both the length of time since population
52 establishment and morphology. Consequently, acroporid communities in the Kushimoto
53 area are likely maintained by survival and growth of existing colonies and/or
54 fragmentation, indicating that conservation of established corals should be the first
55 priority to ensure persistence of coral assemblages in such temperate non-reefal areas.

56

57

58 **Introduction**

59 Coral assemblages in higher-latitude areas, such as subtropical reefs and temperate
60 non-reefal areas, have attracted enormous attention as potentially important refugia for
61 tropical coral species due to global climate change (Riegl 2003; Riegl and Piller 2003;
62 Beger et al. 2014; Yamano et al. 2011; Baird et al. 2012). Abnormally high summer
63 seawater temperatures have induced coral bleaching events, resulting in substantial
64 mortality and structural shifts in tropical coral communities (Hughes et al. 2018, 2019).
65 Effects of thermal bleaching have been comparatively less severe in many higher-latitude
66 coral communities (Hughes et al. 2018). In addition, rising seawater temperatures are
67 gradually extending the distributional limits of coral species to higher latitudes (Precht
68 and Aronson 2004; Greenstein and Pandolfi 2008; Yamano et al. 2011; Baird et al. 2012;
69 Nakamura and Yokochi 2020). In the Solitary Islands (30° S), Australia, four tropical
70 *Acropora* species have recently been observed for the first time (Baird et al. 2012). In
71 Japan, poleward expansion of distributional ranges of tropical coral species to temperate
72 non-reefal areas has been confirmed, based on records since the 1930's (Yamano et al.
73 2011). The lesser impact of thermal bleaching and northward range expansion of tropical
74 corals suggest that subtropical reef and temperate non-reefal regions may serve as refugia
75 in this era of global warming (Riegl 2003; Riegl and Piller 2003; Beger et al. 2014;
76 Nakabayashi et al. 2019); hence, coral communities in those regions need to be given
77 high conservation priority. For this reason, understanding fundamental mechanisms of
78 coral community maintenance and persistence is now urgently required in subtropical reef
79 and temperate non-reefal areas.

80 Recruitment of new individuals is critical for persistence and maintenance of coral
81 communities (Underwood and Fairweather 1989). However, current knowledge of
82 recruitment processes is very limited in subtropical reef and temperate non-reefal areas,
83 and most previous studies were conducted for less than ten years (Table 1). Since
84 recruitment demonstrates spatiotemporal variability due to complex biological and
85 environmental factors (Adjeroud et al. 2017), longer-term data must be gathered to
86 estimate effects of recruitment for community maintenance in these higher-latitude coral
87 populations.

88 The Kushimoto area of Wakayama Prefecture in Japan supports relatively higher-
89 latitude coral habitat (33° N). These populations are located near the northern limit of
90 coral distribution, and these coral assemblages have maintained relatively high species
91 diversity and high coverage for at least 100 years (Sugiyama 1937; Uchinomi 1966;
92 Marine Parks Center 1970; Nomura et al. 2008; Nomura 2009). Today, they comprise

93 roughly 115 reef coral species (Nomura et al. 2016), and coral cover has remained
94 above 30% on average (Biodiversity Center of Japan, 2019), even though coral
95 communities in the area have experienced severe disturbances, such as large typhoons
96 and predation by rock snails (*Drupella fragum*) and crown-of-thorns starfish
97 (*Achantaster planci*) (Nomura 2009). For these reasons, the area was designated as the
98 first national marine park in Japan in 1970, and efforts have been made to conserve
99 marine life and the environment. These results have been internationally evaluated and
100 the area was designated as a registered wetland under the Ramsar Convention in 2005.

101 Recent genetic studies and community surveys have suggested possible recruitment
102 patterns of acroporid corals in the Kushimoto area. *Acropora hyacinthus* (Dana 1846) is
103 found throughout the Indo-Pacific, including temperate non-reefal regions, such as
104 Kushimoto (Veron 2000), where it has been recorded since 1931 (Sugiyama 1937).
105 Nonetheless, the major genetic lineage in temperate non-reefal populations is distinct
106 from those in sub-tropical populations in Japan (Suzuki et al. 2016; Nakabayashi et al.
107 2019). In contrast, *A. muricata* (Linnaeus 1758) has only been recorded in Kushimoto
108 since 1995, where it has since replaced *A. hyacinthus* throughout much of its preferred
109 habitat (Nomura 2009). These results imply that temperate non-reefal populations of *A.*
110 *hyacinthus* have been locally maintained since its establishment, whereas larval supply
111 of *A. muricata* from subtropical areas occurs somehow in the Kushimoto area. However,
112 sexual recruitment patterns in the Kushimoto area have not been confirmed and quantified
113 using settlement panels or other means.

114 In this study, we observed settlement from 2004–2016 (except 2007) in the Kushimoto
115 area to quantify larval supply in these temperate coral populations. We also assessed
116 genetic population structures of these two acroporid species.

117

118 **Methods**

119 **1. Study site**

120 We selected three sites in the Kushimoto area, two along the coast of Kushimoto (St 1
121 and St 2) and another at Cape Shionomisaki (St 3, Fig 1). Amounts and composition of
122 settlement were investigated at these sites. In addition, genetic diversity of both acroporid
123 species at St 1 and St 2 was investigated.

124

125 **2. Settlement surveys**

126 Assessments of settlement were conducted since 2004 at St 1 (except 2007), and 2008
127 at St 2 and St 3. To assess settlement patterns, 8 to 10 artificial settlement panel pairs (Fig

128 2) were deployed at each site at a depth of approximately 5 m. This was done about one
129 month before the predicted major coral spawning periods, so as to pre-condition the
130 panels. In the Kushimoto area, acroporid communities are reproductively active, as
131 evidenced by observations of spawning from 2003 to 2015. Acroporid corals at
132 Kushimoto spawned from the beginning of June to the middle of August (Table 2).

133 Artificial settlement panel pairs consisted of a pair of $10 \times 10 \times 0.6$ cm panels made
134 of fiber-reinforced cement. Two panels were fastened one above the other with a 2-cm
135 separation, so that coral larvae can settle between them and most grazers cannot pass
136 between them. Panel pairs were set haphazardly on substrates, at least 1 m apart, using
137 underwater epoxy glue. They were retrieved from September to November, at least 1
138 month after observed spawning of acroporids (Supplementary Table 1). Retrieved panel
139 pairs were bleached with a chlorine solution for one week to eliminate organic matter and
140 then dried for observation under a stereomicroscope. Upper and lower surfaces of both
141 panels were observed; thus, 0.04 m^2 of surface were sampled for each panel pair (no
142 recruits were observed on the sides of the panels). Coral recruits on panels were identified
143 to the family level (Acroporidae, Poritidae, Pocilloporidae, and others) and counted,
144 based on skeletal morphology (Babcock et al. 2003). Annual variations in mean
145 settlement (mean number of recruits per panel pair) of total corals and of each coral family
146 were analyzed using the Kruskal-Wallis test. Results of settlement were compared with
147 percent cover data collected as part of the Monitoring Sites 1000 Project by the Ministry
148 of Environment and the Biodiversity Centre (<http://www.biodic.go.jp/moni1000/>, in
149 Japanese). Percent cover was estimated using the spot-check method. Changes in % cover
150 at all three sites for 2004 to 2016 are shown in Supplementary Fig 1. To estimate effects
151 of local stocks on settlement, we analyzed whether mean coral settlement of all corals
152 was related to mean % cover at the three sites, using Spearman's rank-order correlation.
153 All analyses were performed in R, version 4.1.0 (R Core Team 2021).

154

155 **3. Genetic diversity**

156 Genetic population structure of the two most abundant coral species in the
157 Kushimoto area, *Acropora hyacinthus* and *A. muricata*, was analyzed to estimate
158 potential sexual recruitment. Forty-three samples of *A. hyacinthus* were collected at 5-m
159 intervals along 50 m of the shore and 15 m from the shore to the near edge of the coral
160 communities. Thirty samples of *A. muricata* were collected every 2 m from a patch of
161 ≥ 16 m along the shore and ≥ 8 m perpendicular to the shore. Specimens were preserved
162 in 99.5% ethanol, and DNA was extracted using a DNeasy Blood & Tissue Kit

163 (QIAGEN) following the standard protocol. Extracted DNA was amplified using
164 multiplex PCR, and four primer sets were added to each PCR tube. Multiplex PCR was
165 performed using a Multiplex PCR Kit (QIAGEN) in a total reaction volume of 10 μ L
166 containing about 50 ng of template genomic DNA, 2 \times Multiplex PCR Master Mix, and
167 0.2 μ M (final concentration) of each of three primers for each locus: a forward primer, a
168 reverse primer with a U19, M13RV, T7, or SP6 tail (*A. muricata*: 846m3/U19,
169 11401m4/M13RV, 441m6/U19, Am01^h/U19, Am02^h/M13RV, Am03^h/T7, Am04^h/SP6,
170 Am05^h/U19, Am06^h/T7, Am07^h/SP6 from Shinzato et al. 2014 and Goossens 2015; *A.*
171 *hyacinthus*: 8346m6/U19, 11401m4/M13RV, Ac0753/U19, Ac0808/T7,
172 Amil2_002/SP6, Amil_006/U19, Amil2_022/M13RV, Amil2_023/T7 from van Oppen
173 et al. 2007, Concepcion et al. 2010, and Shinzato et al. 2014) labeled with FAM, VIC,
174 NED, or PET, respectively. PCR cycling conditions were 15 min at 95 °C, followed by
175 30 cycles of 30 s at 94 °C, 90 s at 57 °C, and 60 s at 72 °C, with an extension of 30 min
176 at 60 °C in the final cycle. When amplification was insufficient, Ampli Taq Gold 360
177 Master Mix (Thermo Fisher Scientific) was used with the following conditions: 95 °C
178 for 9 min followed by 35 cycles at 95 °C for 30 s, 54 °C for 30 s, 72 °C for 1 min, and a
179 final extension of 5 min at 72 °C. Allelic variations of amplified products were analyzed
180 using a DNA capillary sequencer (3130xl Genetic Analyzer, Thermo Fisher Scientific)
181 and GeneMapper ver. 3.7 (Thermo Fisher Scientific). GenClone ver. 2.0 (Arnaud-Haond
182 and Belkhir 2007) was used to determine clonality in the populations. Clonal replicates
183 were removed according to the results of GeneClone ver.2.0, and GenAlEx ver.6.5
184 (Peakall and Smouse 2006) was used to calculate the probability of identity (PI),
185 number of alleles (Na), observed and expected heterozygosities (Ho and He,
186 respectively), and deviation from Hardy-Weinberg equilibrium (F).

187

188 **Results**

189 In all, 1086 coral recruits were observed on 278 settlement panel pairs for 2004 – 2016
190 (no data in 2007, Table 3). Total settlement was dominated by the family Poritidae, which
191 accounted for 83.0% (902 recruits) of total settlement. Acroporid and pocilloporid
192 settlements comprised only 6.0% (66 recruits) and 10.9% (119 recruits) of total settlement,
193 respectively.

194 Each year, >75% of all settlement panel pairs recorded 0-9 coral recruits per panel pair
195 (Fig 3). Notably, for the family Acroporidae, more than 80% of all settlement panel pairs
196 of each year recorded 0 recruits. In 2008 – 2014 and 2016, one to three acroporid recruits
197 per panel pair were observed, except in 2014. In 2014, 29 acroporid recruits were

198 observed in total and one of 25 panel pairs recorded 11 recruits (Table 3, Fig 3). Mean
199 acroporid recruits per panel pair were fewer than 1 in most years (1.16 ± 0.53 , mean
200 \pm SE) in 2014 (Fig 4). For the family Pocilloporidae, proportions of panel pairs with 0
201 and <10 recruits per year were 69.2-100% and 0-42.3%, respectively. However, 21
202 recruits were observed on a panel pair in 2012. As for acroporids, mean pocilloporid
203 recruits per panel pair were <1 most years, but 1.92 ± 0.87 in 2012. Comparatively, the
204 proportion of panels with >10 recruits per pair was 3.5-23.3% (for 2008 – 2014 and 2016)
205 for the family Poritidae. In 2008, 195 poritid recruits were observed on one panel pair.
206 Relatively, mean poritid recruits per panel pair were >1 in most years with 14.0 ± 6.7
207 in 2008. There was no correlation between mean settlement and mean % cover of total
208 corals ($r = -0.02$, Spearman's rank correlation, Supplementary Fig 2).

209 Thirty samples taken from a patch of *A. muricata* showed the same genotypes for all
210 10 loci and were therefore considered clones. However, for *A. hyacinthus*, we identified
211 41 genotypes, with 87.5% polymorphic loci, among the 43 samples. There were two
212 clonal groups in *A. hyacinthus* populations, and clonemates within each group were
213 approximately 5 m apart. With multilocus genotypes ($G=41$), the probability of identity
214 (PI) at the eight loci was 2.4×10^{-4} . The number of alleles ranged from one to nine for
215 eight loci (average 5.1 per locus; Table 4). Mean observed and expected heterozygosities
216 were 0.21 ± 0.08 and 0.39 ± 0.09 , respectively. Deviation from Hardy-Weinberg
217 equilibrium ranged from 0.168 to 1.000 for the eight loci.

218

219 **Discussion**

220 Recruitment of acroporid corals was low, even though acroporid corals are one of the
221 foundation species in the area. Pocilloporid recruits were also few in number, but poritid
222 corals showed relatively high recruitment and dominated total recruitment in the
223 Kushimoto area. The dominance of poritid corals contrasted with results of similar studies
224 conducted in subtropical reefs and temperate non-reefal areas ($>26^\circ$ N/S latitude), at
225 which pocilloporids tended to dominate recruitment on settlement panels (Table 1).
226 Earlier studies also reported a scarcity of acroporid recruits (Harriott & Banks 1995;
227 Harriott 1999; Nozawa et al. 2006). In the Solitary Islands, low acroporid recruitment has
228 been consistently recorded (Harriott 1999), while coral cover of the area has been stable
229 during the past 10 to 15 years (Dalton & Roff 2013). These results suggest that relatively
230 low larval supply may be sufficient to maintain the populations. Similarly, coral
231 assemblages in the Kushimoto area, mainly composed of acroporid corals, have
232 maintained relatively high cover for over a decade. Coral cover remained >20%, and

233 recently increased more than 30% in the area (Supplementary Fig. 1). This is relatively
234 high coverage compared with that in other regions worldwide, with most showing <25%
235 cover from 1997 to 2004 (Bruno and Selig 2007). The relatively high coral cover in the
236 Kushimoto area may have resulted from asexual growth and low mortality of existing
237 corals because acroporid recruits in the area were 0 or fewer than 1 per panel pair in most
238 years. This suggests that sexual recruitment could be of limited importance for
239 community maintenance in the Kushimoto area. On the other hand, it also implies that
240 relatively low recruitment may be sufficient for community maintenance.

241 Constant low recruitment suggests the possibility that new genotypes are rarely
242 added to the population at Kushimoto. In addition, recently established populations tend
243 to have relatively lower clonal diversity (Nakabayashi et al. 2019). Therefore, instead of
244 the potential for rare recruitment of new genotypes, differences in the length of time
245 since population establishment may affect genetic structure of populations in this area.
246 The tabular acroporid, *A. hyacinthus*, is widely distributed from subtropical reefs to
247 temperate non-reefal areas in Japan (Suzuki et al. 2016; Nakabayashi et al. 2019;
248 Nakamura and Yokochi 2020), including the Kushimoto area, where this acroporid
249 species has been recorded since 1931 (Sugiyama 1937). That is, this population has
250 been maintained for nearly 100 years. *A. hyacinthus* is a long-term resident in the area.
251 Over the years, new recruits, even though few in number, may have intermittently
252 settled in the area, resulting in relatively higher genotypic diversity compared to the
253 recently arrived species, *A. muricata*. A branching acroporid, *A. muricata* was first
254 observed at Kushimoto in 1995. In Kushimoto, the annual average seawater temperature
255 has risen abruptly since the 1990s (Fig 5), and colonization of some tropical coral
256 species that had not been seen before, including *A. muricata*, has been observed. After
257 colonization, *A. muricata* began to invade areas covered by *A. hyacinthus* (Nomura
258 2009). Percent coverage of *A. muricata* was only ~5% in 1995, in comparison with
259 approximately 70% for *A. hyacinthus*, but by 2002, both species showed coverage of
260 ~40%, and by the following year, *A. muricata* had achieved approximately 60%
261 coverage. In October 2004, the *A. muricata* population was reduced by a typhoon, and
262 many colonies broke into fragments due to their arborescent growth form. However, re-
263 growth of remnants and fragments was observed in subsequent years (Nomura 2009).
264 That may explain why the observed patch of *A. muricata* consists of only a single
265 genotype. On the other hand, due to limited acroporid recruitment, recently arrived *A.*
266 *muricata* may not have had enough time to establish patches with different genotypes;

267 however, sampling from other patches of *A. muricata* will be necessary to better
268 estimate the genetic structure of the population.

269 When sexual recruitment occurs at low levels, asexual reproduction by clonal growth
270 and fragmentation may allow population persistence. Recovery due to asexual
271 reproduction by fragmentation and reattachment of branches has been observed in
272 branching acroporid corals after hurricanes in the Caribbean (Highsmith 1982). However,
273 while tabular acroporids are considered more vulnerable to physical disturbances, e.g.,
274 wave action during storms or hurricanes, similar to branching corals (Muko et al. 2013),
275 fragments of tabular corals are less likely to survive, because of higher risk of polyp
276 disorientation (Smith and Hughes 1999). In addition, as tabular *A. hyacinthus* at
277 Kushimoto have relatively short stalks and tend to grow close to the substrate (Fig 6),
278 storm-induced fragmentation of tabular *A. hyacinthus* at Kushimoto is less severe than
279 might be expected. This could be one of the reasons that the *A. hyacinthus* population
280 shows less clonality than *A. muricata* in the Kushimoto area.

281 Our findings imply that acroporid communities in the Kushimoto area are likely
282 maintained by survival and growth of existing colonies and/or fragmentation. Even
283 though acroporid corals are reproductively active (Misaki 2017), gametes produced
284 locally may largely be swept away by currents because of the lack of reefs; therefore,
285 sexual recruitment has been low for over a decade. From these findings, if coral
286 assemblages in the Kushimoto area become degraded due to drastic environmental
287 changes, they are unlikely to be resupplied by larvae from either the local area or the
288 upper stream of the Kuroshio Current. Their recovery would depend mainly on asexual
289 growth of remnant colonies.

290 As a designated Wetland of International Importance under the Ramsar Convention,
291 specifically because of coral assemblages in the Kushimoto area, conservation of existing
292 corals should be a top priority for sustainability of marine ecosystems in the area.
293 Moreover, a recent taxonomic study in higher-latitude areas revealed a number of
294 unidentified coral species in the area, suggesting that higher-latitude coral communities
295 may contain a number of unidentified endemic species (Nomura et al. 2020). Because a
296 low level of recruitment is observed at Kushimoto and also in other higher-latitude coral
297 assemblages (Harriott & Banks 1995; Harriott 1999; Nozawa et al. 2006), conservation
298 measures for existing corals should be the first priority to ensure persistence of coral
299 assemblages in these areas, indicating that a re-evaluation of conservation strategies in
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314 **References**

- 315 Adjeroud M, Kayal M, Penin L (2017) Importance of recruitment processes in the
316 dynamics and resilience of coral reef assemblages. *Marine Animal Forests*, 549-569.
- 317 Arnaud-Haond S, Belkhir K (2007) Geneclone: a computer program to analyse genotypic
318 data, test for clonality and describe spatial clonal organization. *Mol Ecol Notes* 7:15-
319 17
- 320 Babcock RC, Baird AH, Piromvaragorn S, Thomson DP, Willis BL (2003) Identification
321 of scleractinian coral recruits from Indo-Pacific reefs. *Zool Stud* 42:211-226
- 322 Baird AH, Sommer B, Madin JS (2012) Pole-ward range expansion of *Acropora* spp.
323 along the east coast of Australia. *Coral Reefs* 31:1063
- 324 Begger M, Sommer B, Harrison PL, Smith SDA, Pandolfi JM (2014) Conserving potential
325 coral reef refuges at high latitudes. *Divers Distrib* 20:245-257
- 326 Biodiversity Center of Japan (2019) Preliminary report on monitoring 1000 coral reef
327 survey in fiscal year 2018. The Ministry of the Environment, Japan.
- 328 Bruno JF, Selig ER (2007) Regional decline of coral cover in the Indo-Pacific: timing,
329 extent and subregional comparisons. *PLoS ONE* 2(8):e711
- 330 Concepcion GT, Polato NR, Baums IB, Toonen RJ (2010) Development of microsatellite
331 markers from four Hawaiian corals: *Acropora cytherea*, *Fungia scutaria*, *Montipora*
332 *capitata* and *Porites lobata*. *Conserv Genet Resour* 2:11-15
- 333 Glasson D, Celliers L, Schleyer MH (2006) Coral recruitment patterns at Sodwana Bay,
334 South Africa. *Coral Reefs* 25:485-492
- 335 Goossens B (2015) Microsatellite records. *Conserv Genet Resour* 7:917-944

336 Greenstein BJ, Pandolfi JM (2008) Escaping the heat: range shifts of reef coral taxa in
337 coastal Western Australia. *Glob Chang Biol* 14:513-528

338 Harriott VJ (1992) Recruitment patterns of scleractinian corals in an isolated sub-tropical
339 reef system. *Coral Reefs* 11:215-219

340 Harriott VJ (1999) Coral recruitment at a high latitude Pacific site: a comparison with
341 Atlantic reefs. *Bull Mar Sci* 65:881-891

342 Harriott VJ, Banks SA (1995) Recruitment of scleractinian corals in the Solitary Islands
343 Marine Reserve, a high latitude coral-dominated community in Eastern Australia. *Mar*
344 *Ecol Prog Ser* 123:155-161

345 Highsmith RC (1982) Reproduction by fragmentation in corals. *Mar Ecol Prog Ser* 7:207-
346 226

347 Hughes TP, Anderson KD, Connolly SR, Heron SF, Kerry JT, Lough JM, Baird AH,
348 Baum JK, Berumen ML, Bridge TC, Claar DC, Eakin CM, Glimour JP, Graham NAJ,
349 Harrison H, Hoobs J-P A, Hoey AS, Hoogenboom M, Lowe RJ, McCulloch MT,
350 Pandolfi JM, Pratchett MS, Schoepf V, Torda G, Wilson SK (2018) Spatial and
351 temporal patterns of mass bleaching of corals in the Anthropocene. *Science* 359:80-83

352 Hughes TP, Kerry JT, Baird AH, Connolly SR, Chase TJ, Dietzel A, Hill T, Hoey AS,
353 Hoogenboom MO, Jacobson M, Kerwell A, Madin JS, Mieog A, Paley AS, Pratchett
354 MS, Torda G, Woods RM (2019) Global warming impairs stoch-recruitment dynamics
355 of corals. *Nature* 568:387-390

356 Marine Parks Center (1970) Overview of the first designated National Marine Park
357 Areas. *Marine Parks Journal* (in Japanese) 93: 8-13

358 Markey KL, Abdo DA, Evans SN, Bosserelle C (2016) Keeping it local: dispersal
359 limitations of coral larvae to the high latitude coral reefs of the Houtman Abrolhos
360 Islands. *PLoS ONE* 11(1):e0147628

361 Misaki H (2017) Spawning patterns of high latitude scleractinian corals from 1989 to
362 2012 at Kushimoto, Wakayama, Japan. *Nankiseibutsu* (in Japanese) 59:55-60

363 Muko S, Arakaki S, Nagao M, Sakai K (2013) Growth form-dependent response to
364 physical disturbance and thermal stress in *Acropora* corals. *Coral reefs* 32:269-280

365 Nakabayashi A, Yamakita T, Nakamura T, Aizawa H, Kitano YF, Iguchi A, Yamano H,
366 Nagai S, Agostini S, Teshima KM, Yasuda N (2019) The potential role of temperate
367 Japanese regions as refugia for the coral *Acropora hyacinthus* in the face of climate
368 change. *Sci Rep* 9:1892

369 Nakamura M, Sakai K (2010) Spatiotemporal variability in recruitment around Iriomote
370 Island, Ryukyu Archipelago, Japan: implications for dispersal of spawning corals. *Mar*

371 Biol 157:801-810

372 Nakamura M, Yokochi H (2020) Updated northern distributional limits of *Acropora* spp.
373 along the Kuroshio Current. *Galaxea, Journal of Coral Reef Studies* 22:3-4

374 Nakamura M, Kumagai NH, Sakai K, Okaji K, Ogasawara K, Mitarai S (2015) Spatial
375 variability in recruitment of acroporid corals and predatory starfish along the Onna
376 coast, Okinawa, Japan. *Mar Ecol Prog Ser* 540:1-12

377 Nomura K (2009) Recent changes in coral communities in Kushimoto, the southernmost
378 part of Honshu, Japan (in Japanese). *Journal of the Japanese Coral Reef Society* 11:39-
379 49

380 Nomura K, Uchida H, Fukuda T (2008) Progressive faunal inventory of hermatypic corals
381 in Kushimoto, Wakayama Prefecture, Japan. *Nankiseibutu (in Japanese)* 50: 191-200

382 Nomura K, Fukami H, Zayasu Y, Shimada G, Kitano Y, Yokochi H, Shimoike K,
383 Tachikawa H, Oku Y, Suzuki G, Kajiwara K (2016) Revision of the zooxanthellate
384 scleractinian corals in Kushimoto, Wakayama, Japan (in Japanese). *Marine Pavilion,*
385 *Supplement 4:1-20*

386 Nomura K, Yokochi H, Kimura T, Kajiwara K, Nojima S, Arakaki S (2020)
387 Zooxanthellate scleractinian corals collated from Amakusa, western Kyushu, Japan.
388 *Coastal Ecosystems* 7:1-52

389 Nozawa Y, Tokeshi MT, Nojima S (2006) Reproduction and recruitment of scleractinian
390 corals in a high-latitude coral community, Amakusa, southwestern Japan. *Mar Biol*
391 149:1047-1058

392 Peakall R, Smouse PE (2006) GenAlEx6: genetic analysis in Excel. Population genetic
393 software for teaching and research. *Mol Ecol Notes* 6:288-295

394 Precht WF, Aronson RB (2004) Climate flickers and range shift of reef corals. *Front. Ecol.*
395 *Environ* 2:307-314

396 Riegl B (2003) Climate change and coral reefs: different effects in two high-latitude areas
397 (Arabian Gulf, South Africa). *Coral Reefs* 22:433-446

398 Riegl B, Piller WE (2003) Possible refugia for reefs in times of environmental stress. *Int.*
399 *J Earth Sci* 92:520-531

400 Shinzato C, Yasuoka Y, Mungpakdee S, Arakaki N, Fujie M, Nakajima Y, Satoh N (2014)
401 Development of novel, cross-species microsatellite markers for *Acropora* corals using
402 next-generation sequencing technology. *Front Mar Sci* 1:11

403 Smith LD, Hughes TP (1999) An experimental assessment of survival, re-attachment
404 and fecundity of coral fragments. *J Exp Mar Biol Ecol* 235:147-164

- 405 Sugiyama (1937) On the recent reef-building corals found in the Japanese Seas.
406 Contributions from the Institute of Geology and Paleontology, Tohoku University (in
407 Japanese) 26: 1-60
- 408 Suzuki G, Keshavmurthy S, Hayashibara T, Wallace CC, Shirayama Y, Chen CA, Fukami
409 H (2016) Genetic evidence of peripheral isolation and low diversity in marginal
410 populations of the *Acropora hyacinthus* complex. Coral Reefs 35:1419-1432
- 411 Uchinomi F (1966) Outline of shallow coral fauna in the Kii Peninsula. Report of the
412 national park candidate site in Wakayama Prefecture. Nature Conservation Society of
413 Japan (in Japanese)
- 414 Underwood AJ, Fairweather PG (1989) Supply-side ecology and benthic marine
415 assemblages. TREE 4:16-20
- 416 van Oppen MJH, Underwood JN, Muirhead AN, Peplow Let (2007) Ten microsatellite
417 loci for the reef-building coral *Acropora millepora* (Cnidaria, Scleractinia) from the
418 Great Barrier Reef, Australia. Mol Ecol Notes 7:436-438
- 419 Watanabe M, Iwase F, Yokochi H (2009) Coral recruitment patterns in high-latitude coral
420 communities in southwestern Shikoku, Japan (in Japanese). Journal of the Japanese
421 Coral Reef Society 11:73-81
- 422 Yamano H, Sugihara K, Nomura K (2011) Rapid poleward range expansion of tropical
423 reef corals in response to rising sea surface temperatures. Geophys Res Lett 38:L04601

424 **Table 1.** Summary of coral recruitment at subtropical marginal reefs and temperate non-reefal areas (>26° N/S latitude)

Area	Site	Latitude	Survey periods	Size of settlement panels/ plates/ tiles	Months of deployment	Number of settlement panels/ plates/ tiles	Total recruits during the survey	Mean recruits	Most abundant family/genus of recruits	Ref
South Africa	Sodwana Bay	27 - 28 °S	1999 to 2002	12 × 12 × 1 cm	1 to 16 months	570 panel pairs	2124 recruits	13.2 per tile for the highest	Pocilloporidae	Glasson et al. 2006
Western Australia	Houtman Abrolhos Islands	28 - 29 °S	2011 to 2013	12 × 12 × 1 cm	5 months	135 tiles		0.4 - 128.8 recruits per tile	Acroporidae	Markey et al. 2016
Eastern Australia	Solitary Islands	30 °S	1992 to 1993	15 × 15 cm	3 to 5 months	120 plate pairs	190 recruits	0.0 - 8.0 recruits per plate pair	Pocilloporidae	Harriott & Banks 1995
			1993 to 1998	15 × 15 cm	5 months	570 panel pairs		0.1 - 20.3 per panel pair	Pocilloporidae	Harriott 1999
Eastern Australia	Lord Howe Island	31 °S	1991	15 × 15 cm	2 to 8 months		585 recruits	3.5 - 48.5 per tile pair	Pocilloporidae	Harriott 1992
Japan	Amakusa	32 °N	2001 to 2003	10 × 10 × 0.4 cm	3 months	200 plates	4 recruits		<i>Acropora / Alveopora</i>	Nozawa et al. 2006
Japan	Southwestern Shikoku	32 °N	2007 to 2008	10 × 10 × 0.5 cm	2 months	400 panel pairs		3.2 -18.0 per panel pair	Pocilloporidae	Watanabe et al. 2009

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439 **Table 2.** Summary of spawning patterns of major acroporid corals in the Kushimoto area.

Species	Season of spawning	Days from the full moon		Days from the new moon	
<i>Acropora hyacinthus</i>	The beginning of July to the beginning of August	6.6 ± 2.3	[3~11days]	5.0 ± 1.0	[4 ~ 6 days]
<i>Acropora muricata</i>	The beginning of June to the beginning of July	7.6 ± 1.9 (mean ± SE)	[4~10 days]	13.5 ± 0.7 (mean ± SE)	[13 ~ 14 days]
<i>Acropora solitaryensis</i>	The middle of July to the middle of August	7.5 ± 0.7	[7~8 days]	5.1 ± 1.8	[2 ~ 8 days]

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442 **Table 3.** Total settlement recorded on all settlement panel pairs by family per year. The
 443 total number of coral recruits did not correlate with weeks of deployment of settlement
 444 panel pairs ($r=0.099$, Pearson's correlation).

Year	Weeks of deployment	Number of panel pairs	Total	Acroporidae	Pocilloporidae	Poritidae	Others
2004	15 weeks	9	0	0	0	0	0
2005	12 weeks	8	0	0	0	0	0
2006	12 weeks	8	0	0	2	0	0
2007	—	—	—	—	—	—	—
2008	15 weeks	30	436	5	12	419	0
2009	14 weeks	28	93	8	11	74	0
2010	22 weeks	30	78	9	14	55	0
2011	20 weeks	30	132	2	12	117	1
2012	20weeks	26	147	2	50	95	0
2013	23 weeks	28	58	7	6	45	0
2014	22 weeks	25	88	29	4	55	0
2015	15 weeks	30	8	0	0	8	0
2016	18 weeks	26	46	4	8	34	0
Total		278	1086	66	119	902	1

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Table 4. Population genetic indices for *A. hyacinthus* for 8 loci. Numbers of multilocus genotypes (G), numbers of alleles (Na), observed and expected heterozygosities (Ho and He, respectively), and deviation index from Hardy-Weinberg equilibrium (F). * p <0.001.

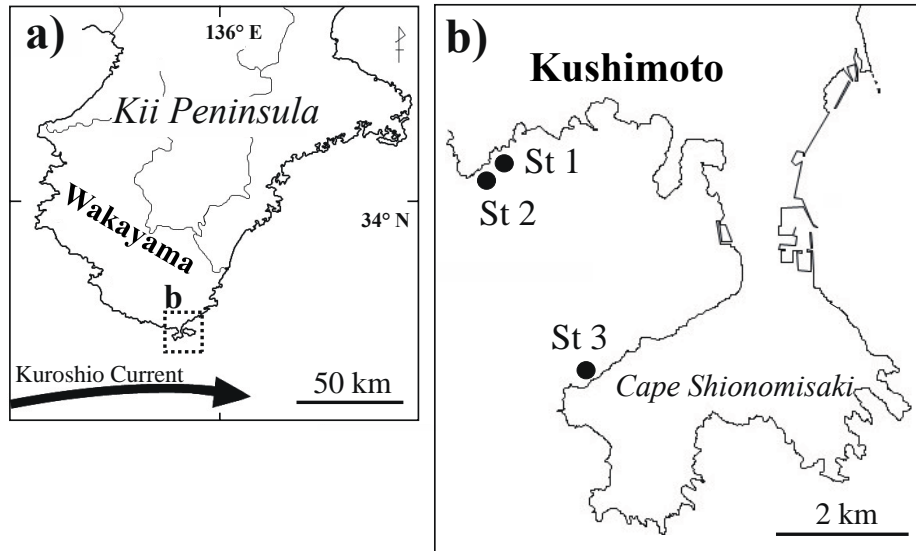
Locus	8346m3	11401m4	Ac0753	Ac0808	Amil2-002	Amil2-006	Amil2-022	Amil2-023	Total (Mean ± SE)
G	41	41	41	41	41	41	41	41	
Na	4	9	2	1	5	7	8	5	5.125 ± 0.990
Ho	0.073	0.610	0.000	0.000	0.293	0.366	0.317	0.000	0.207 ± 0.079
He	0.386	0.802	0.093	0.000	0.389	0.483	0.381	0.621	0.394 ± 0.092
F	0.810*	0.240*	1.000*	—	0.248*	0.243*	0.168*	1.000*	0.530 ± 0.137

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456 **Figures:**



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458 **Fig. 1** Study sites. a) Wakayama Prefecture in the Kii Peninsula, b) the Kushimoto area.

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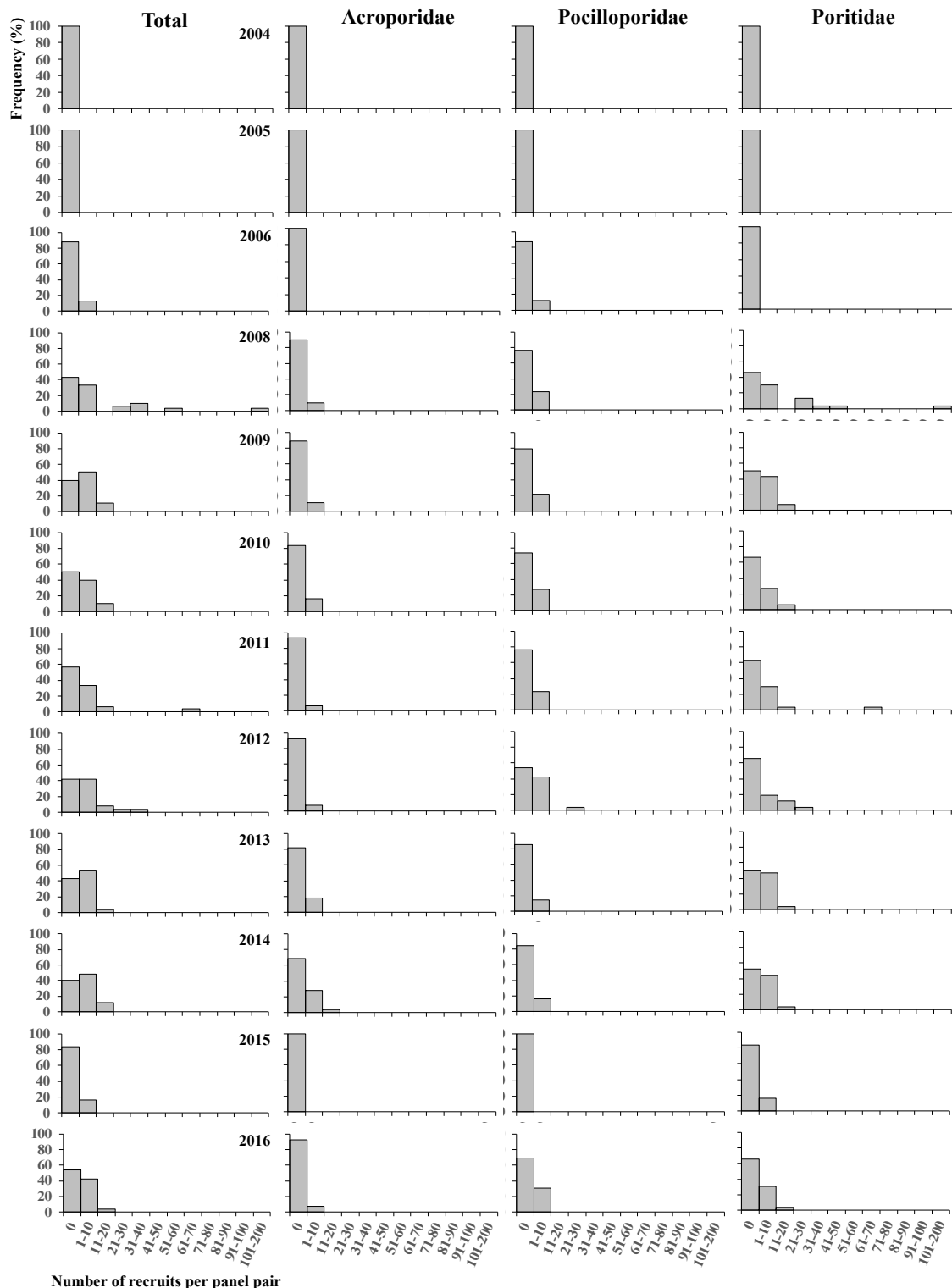
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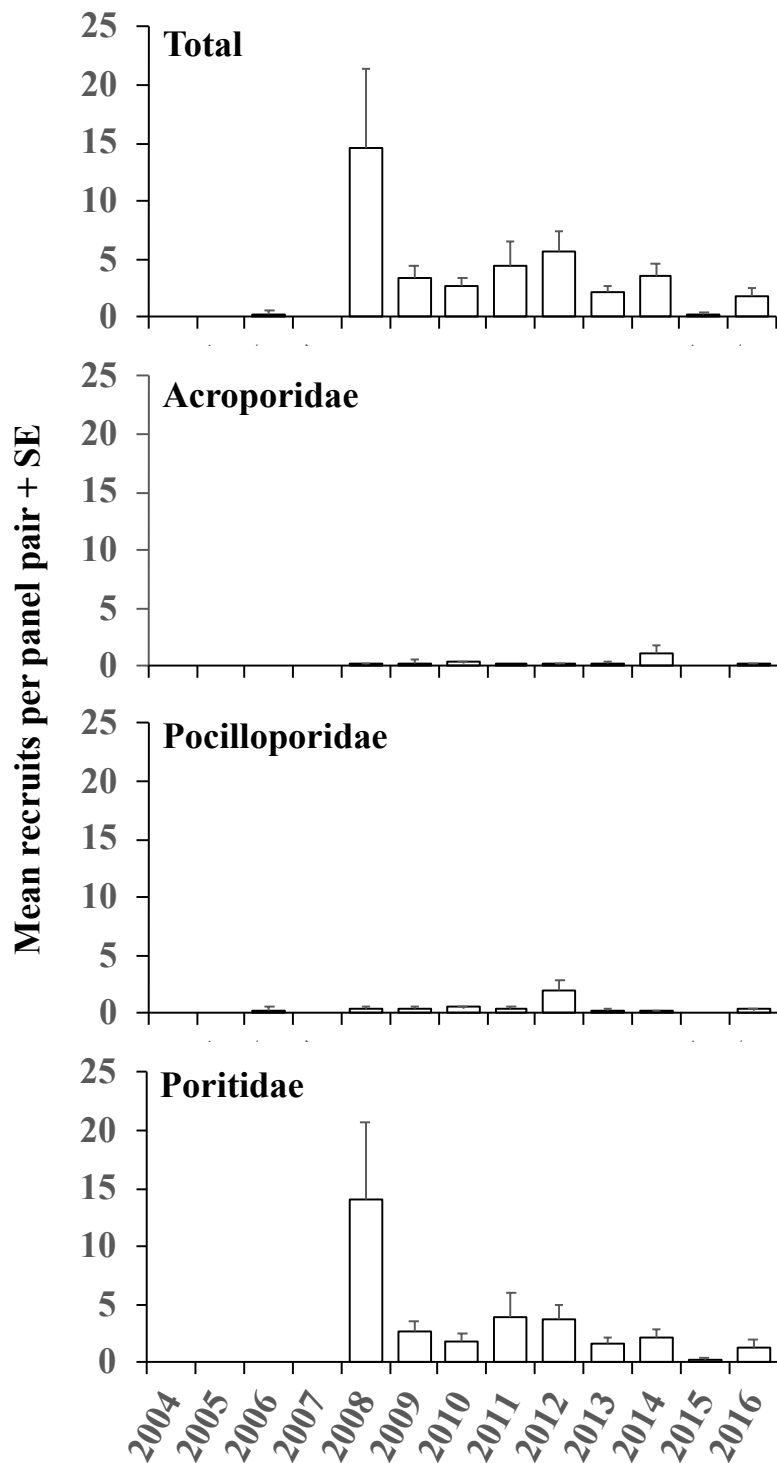
473 **Fig. 2** Settlement panel pairs, just after deployment.

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476 **Fig. 3** Frequency distributions of numbers of total coral recruits, Acroporidae,
 477 Pocilloporidae and Poritidae per panel pair from 2004 to 2016, except 2007 at Kushimoto,
 478 Wakayama Prefecture, Japan. Data of the three sites were pooled.



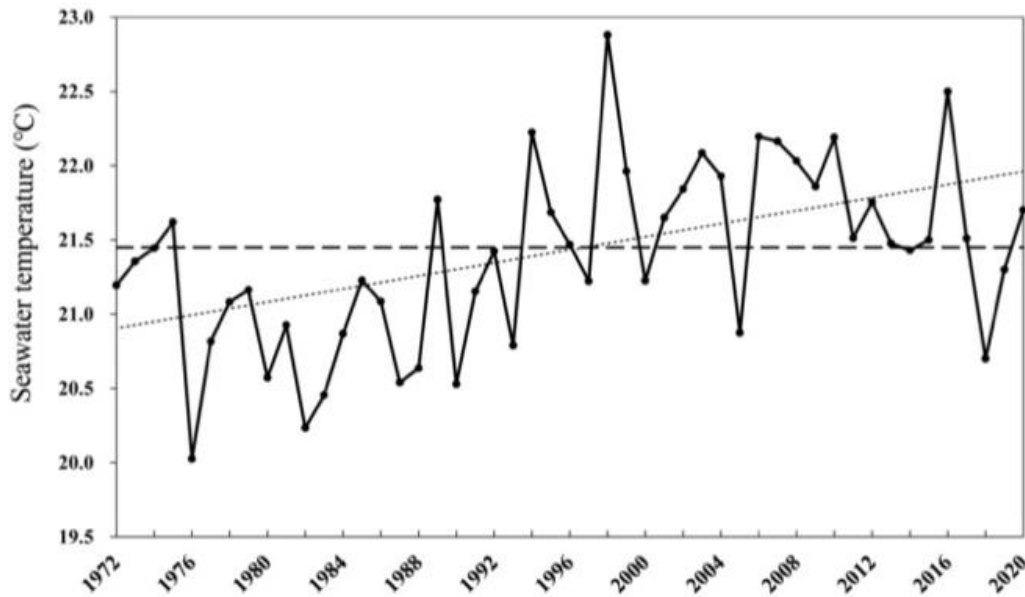
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480 **Fig. 4** Annual variation in settlement for 2004-2016 (except 2007) in the Kushimoto area.

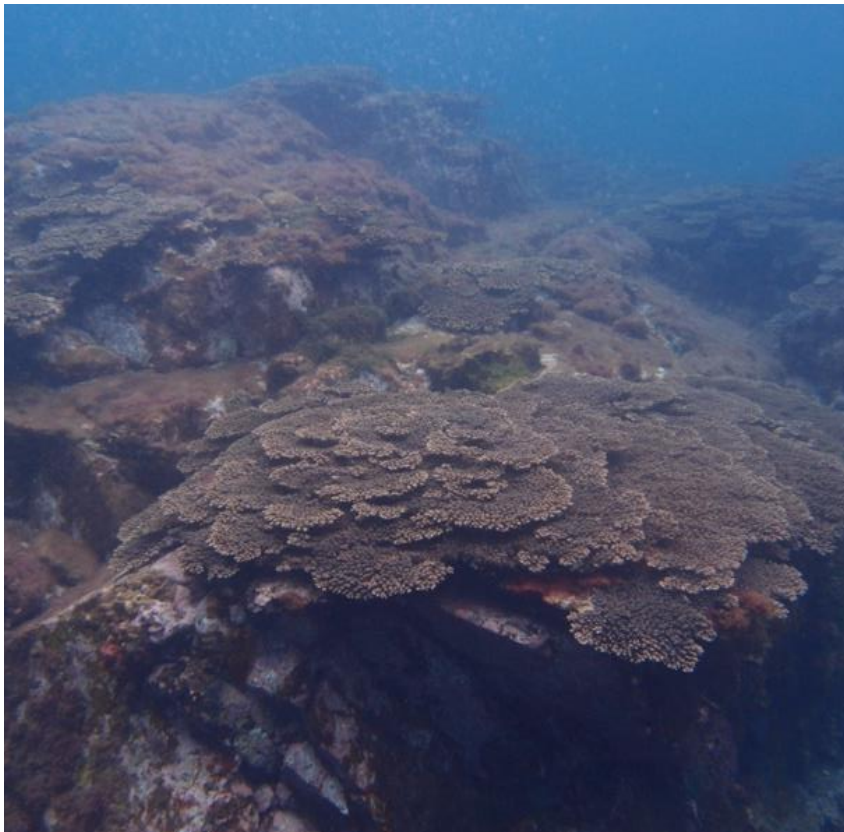
481 Data from three sites were pooled. Mean settlement of all corals and of each coral family

482 differed significantly among years ($p < 0.05$, Kruskal-Wallis test).

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 485 **Fig. 5** Annual average surface water temperature at Kushimoto, Wakayama Prefecture,
 486 Japan. Data were recorded at St 1. The mean annual average surface temperature, from
 487 1981 – 2010, was 21.45°C, an increase of 1.1°C over the last 50 years (dashed line). The
 488 regression line ($y = 0.0209x - 20.2685$, $r=0.491$, $p < 0.05$) is shown as a dotted line.
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 491 **Fig. 6** *Acropora hyacinthus* at Kushimoto, Wakayama Prefecture, Japan.

Year	Date of Deployment	Date of Retrieval
2004	June 3 rd	September 25 th
2005	June 18 th	September 15 th
2006	June 14 th	September 14 th
2007	—	—
2008	June 8 th & 9 th	October 3 rd
2009	May 30 th & 31 st	September 17 th
2010	May 21 st & 22 nd	November 2 nd
2011	May 25 th & 26 th	October 24 th
2012	May 23 rd & 24 th	October 22 nd
2013	May 22 nd & 23 rd	November 11 th
2014	May 28 th & 29 th	November 17 th
2015	June 17 th & 18 th	October 6 th & 7 th
2016	May 14 th & 15 th	September 30 th

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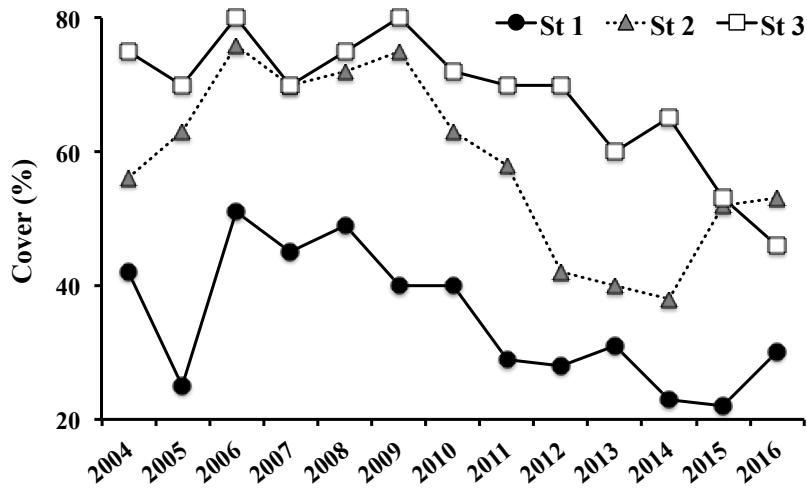
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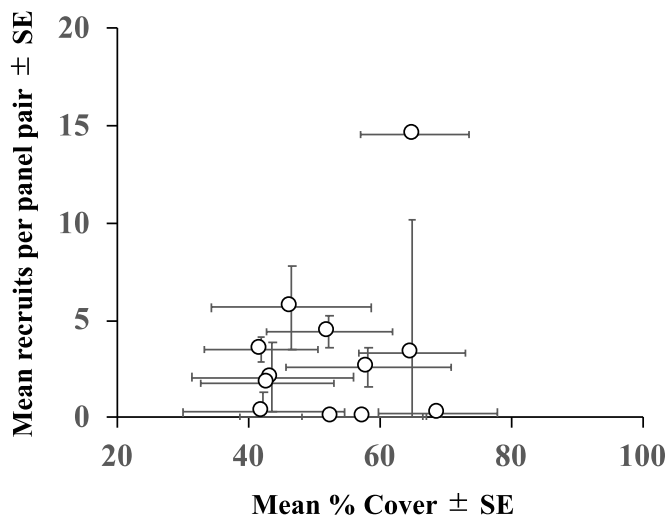
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Supplementary Fig. 1 Coral % cover at three sites from 2004-2016 at Kushimoto, Wakayama Prefecture, Japan, estimated using the Spot Check Method. At St 1 and St 2, percent coral cover essentially equated to acroporid % cover, because coral communities at these sites consisted mainly of *Acropora* spp., especially *A. hyacinthus* and *A. muricata*.



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Supplementary Fig. 2 Relationship between mean settlement and mean percent cover of all corals.